IA-32

Machine learning supported image analysis of microfluidic droplets: Using Random Forest classifiers and Bayesian inference for identification of experimental conditions

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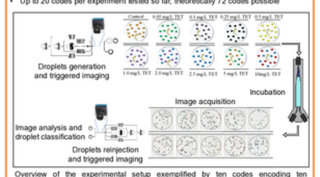
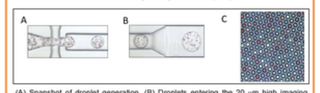
1. Experimental setup

- Coincubation of *Escherichia coli* and antibiotics in 300 pL droplets
- Encoding experimental conditions by combinations of bead colors
- Analysis of droplet triggered images
- Up to 20 codes per experiment tested so far, theoretically 72 codes possible



Overview of the experimental setup exemplified by ten codes encoding ten concentrations of the antibiotic Tetracycline hydrochloride (TET).



(A) Snapshot of droplet generation. (B) Droplets entering the 20 μm high imaging channel. (C) Phase contrast image of droplets in the incubator.

2. Segmentation

Droplet and bead segmentation

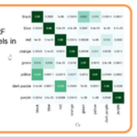
1989 beads: $Acc = 0.979$, $Rec = 0.983$ and $Pre = 0.996$

Growth detection

3000 droplets: $Acc = 0.925$, $Rec = 0.788$ and $Pre = 0.992$

3. Bead classification

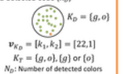
C_T: True bead color
C_D: Bead color detected by RF
RF are resistant to faulty labels in training data [1]



- 8 bead colors
- 11x11 pixels
- Lab colour space
- Random forest (RF) with 5000 trees
- Implemented in Python using *scikit-learn*

4. Droplet decoding

- Each droplet has a true code (K_T) and the RF gives a detected code (K_D)
  $K_D = \{C_{D,1}, \dots, C_{D,N_D}\}$
- We are looking for the most likely code given:
  - Bayes theorem
    $p(K_T|K_D) \sim p(K_D|K_T)p(K_T)$
  - The misclassification rate of the RF
    $p(K_D|K_T) = \prod_{j=1}^{N_D} p(C_{D,j}|C_{T,j})^{k_j}$ ← Number of beads of color j
  - A multinomial distribution with equal probability for each color
    $p(K_T) = p_{MN}(\boldsymbol{v}_{K_T}) = \frac{\sum_{j=1}^{N_T} k_j}{\prod_{j=1}^{N_T}(k_j!)} \left(\frac{1}{N_T}\right)^{\sum_{j=1}^{N_T} k_j}$



$K_D = \{g, o\}$
$\boldsymbol{v}_{K_D} = [k_1, k_2] = [22,1]$
$K_T = \{g, g\}$, $\{g\}$ or $\{o\}$
N_D: Number of detected colors in a droplet

$p(\{g\}) = p(g|g)^{22}p(o|g)^1 p_{MN}([23]) \approx 7.98 \cdot 10^{-5}$ Most probable code
$p(\{o\}) = p(g|o)^{22}p(o|o)^1 p_{MN}([23]) \approx 1.82 \cdot 10^{-76}$
$p(\{g, o\}) = p(g|g)^{22}p(o|o)^1 p_{MN}([22,1]) \approx 1.39 \cdot 10^{-6}$

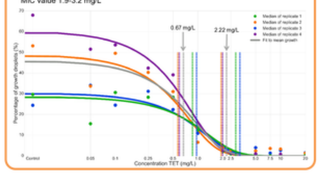
- Adapting to each new dataset
- Automatic re-training of the RF
- Single color droplets identified

	Acc, pre-trained RF	Acc, re-trained RF
Generation	0.94	0.97
8 h incubation	0.92	0.99

- 4259 droplets manually validated
- 18 were wrongly decoded (<0.5%)

5. Minimum inhibitory concentration

- Known MIC of TET versus *E. coli* is ~1-2 mg/L [3]
- Three replicates with 12 concentrations, one with 2x10 concentrations
- Percentage of droplets with growth as antimicrobial susceptibility indication
- Fit $G(\{C\}) = 2A\left(1 - \frac{1}{1+e^{-4[C]}}\right)$ to the growth data, the 50% and 5% levels are used as MIC limits
- In all individual replicates lower MIC value were in range 0.5-1.0 mgL and the upper MIC value 1.9-3.2 mg/L



6. Conclusions

- Method able to encode and decode at least 20 experimental conditions
- No need for fluorescent dyes and laser illumination that may interfere with biological activity
- To be merged with current growth detection and sorting algorithm [3]

References
[1] Svensson *et al.*, (2015) Automated classification of circulating tumor cells and the impact of interobsever variability on classifier training and performance. *Journal of Immunology Research*, ID 573165
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[3] Zhang, Brandes *et al.*, (2013) Real-time image processing for label-free enrichment of Actinobacteria cultivated in picolitre droplets. *Lab on a Chip*, 13(18):3707-13.

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